(DMSO- d_{6} , 300 MHz) δ 1.20 (s, 9 H), 11.07 (s, 1 H), 12.19 (s, 1 H), 13.8 (br s, 1 H). Anal. Calcd for $C_{10}H_{12}BrN_5O_2$: C, 38.23; H, 3.85; N, 22.29; Br, 25.44. Found: C, 38.08; H, 4.04; N, 22.01; Br, 25.68.

Dimethyl [4-[(2-Pivaloylguanin-8-yl)ethynyl]benzoyl]glutamate (7). To a solution of 1.1 g (3.5 mmol) of 5a in 20 mL of CH₃CN and 2 mL of Et₃N was added a mixture of 82 mg (0.36 mmol) of Pd(OAc)₂, 108 mg (0.57 mmol) of CuI, and 213 mg (0.81 mmol) of Ph₃P followed by a solution of 2.02 g (6.6 mmol) of dimethyl [4-ethynylbenzoyl]glutamate (6)⁶ in 20 mL of CH₃CN. The resulting solution was heated at 70-80 °C for 6 h and then concentrated under reduced pressure, and the resulting solid was purified by flash chromatography with 145 g of silica gel, using 3% MeOH in CH_2Cl_2 as the eluting solvent, to give 0.91 g of a solid which (by NMR) consisted of 60% of 2-pivaloyl-8-bromoguanine 5a and 40% of 7: ¹H NMR (DMSO- d_6 , 300 MHz) δ 1.22 (s, 9 H), 2.0 (m, 1 H), 2.1 (m, 1 H), 2.48 (t, 2 H, J = 7.3 Hz, CH_2CH_2CH), 3.57 (s, 3 H, CH_3), 3.64 (s, 3 H, CH_3), 4.45 (m, 1 H, $CHCH_{2}$, 7.74 (d, 2 H, J = 8.0 Hz, $C_{6}H_{4}$), 7.96 (d, 2 H, J = 8.0Hz, C_6H_4), 8.92 (d, 1 H, J = 7.3 Hz, NHCH), 11.09 (s, 1 H), 12.21 (s, 1 H), 13.68 (br s, 1 H, 9-NH).

Dimethyl [4-[2-(2-Pivaloylguanin-8-yl)ethyl]benzoyl]glutamate (8). A suspension of 0.5 g of 3% palladium-on-charcoal and 0.7 g (1.30 mmol) of the above mixture of 5b and 7 in 40 mL of MeOH was stirred at rt under 50 psi of hydrogen for 16 h. The catalyst was removed by filtration through a pad of Celite, which was washed with 25 mL of 5% MeOH in CH₂Cl₂. The filtrate was concentrated to give a solid which was dissolved in 5% MeOH in CH₂Cl₂. Filtration removed a small amount of a white solid (2-pivaloylguanine), and concentration of the filtrate then gave 0.58 g of an orange solid which was purified by flash chromatography (5% MeOH in CH₂Cl₂) through 86 g of silica gel, yield 0.22 g (31%) of 8 as a yellow solid. The analytical sample, mp 175-176 °C, was prepared by recrystallization from ethyl acetate: IR (KBr) 3140, 2940, 1735, 1660, 1400, 1155 cm⁻¹. For the major tautomer: ¹H NMR (DMSO- d_6 , 300 MHz) δ 1.22 (s, 9 H), 2.0 and 2.1 (2m, 2 H, CH_2CH), 2.42 (t, 2 H, J = 7.3 H, CH_2CH_2CH), 3.06 (m, 4 H, CH₂CH₂), 3.55 (s, 3 H, CH₃), 3.61 (s, 3 H, CH₃), 4.41 (m, 1 H, CHNH), 7.29 (d, 2 H, J = 8.0 Hz, C₆H₄), 7.75 (d, 2 H, J =8.0 Hz, C_6H_4), 8.66 (d, 1 H, J = 7.3 Hz, CHNH), 11.02 (s, 1 H), 12.16 (s, 1 H), 13.08 (s, 1 H). For the minor tautomer, the upper field region (lower than 7 ppm) is the same as that of the major tautomer: ¹H NMR (DMSO- d_6 , 300 MHz) δ 7.31 (d, 2 H, J = 6.1 Hz, C_6H_4), 7.77 (d, 2 H, J = 6.1 Hz, C_6H_4), 8.67 (d, 2 H, J = 7.0 Hz, CHNH), 10.93 (s, 1 H), 12.03 (s, 1 H), 12.60 (s, 1 H). MS (FAB, m/e), 541 (MH⁺), 366; exact mass (FAB) calcd for C₂₆-H₃₃N₆O₇ (MH⁺) 541.2410, found 541.24049. Anal. Calcd for C₂₆H₃₂N₆O₇: C, 57.77; H, 5.97; N, 15.55. Found: C, 57.56; H, 5.78; N. 15.32

[4-(2-Guanin-8-ylethyl)benzoyl]glutamic Acid (2). A suspension of 0.14 g (0.26 mmol) of the ester 8 in 2.5 mL of 1 N NaOH was stirred at rt for 3 days. The resulting clear solution was neutralized with acetic acid until pH 6 and then diluted with 10 mL of water, and the resulting solid was collected by filtration, washed throughly with water, MeOH, and ether, and finally dried to give 50 mg (48%) of a yellow solid, mp 200-230 °C dec: IR (KBr) 3350 (br), 1690, 1630 cm⁻¹; ¹H NMR (DMSO-d₆, 300 MHz) δ 1.95 and 2.05 (2m, 2 H, CH₂CH), 2.32 (s, 2 H, CH₂CH₂CH), 2.9 and 3.34 (2 br s, 4 H, CH₂CH₂), 4.37 (m, 1 H, CHNH), 6.23 (s, 2 H, NH₂), 7.29 (d, 2 H), 7.70 (d, 2 H), 8.51 (d, 1 H, NHCH), 10.5 (s, 1 H), 12.1 (s, 1 H), 12.5 (s, 2 H, 2 COOH); mass (FAB, m/e), 429.5 (MH⁺) 282.4; exact mass (FAB) calcd for C₁₉H₂₁N₆O₆ (MH⁺) 429.1522, found 429.15107. Anal. Calcd for C₁₉H₂₀N₆O₆^{-1/}₂H₂O: C, 52.17; H, 4.84; N, 19.21. Found: C, 52.44; H, 4.60; N, 18.98.

Acknowledgment. We are grateful to the National Institutes of Health (Grant No. CA42367) and to Eli Lilly & Company, Indianapolis, IN for support of this work, to Dr. G. B. Grindey, Eli Lilly & Company, for carrying out the preliminary biological screening for in vitro cytotoxic activity, and to the Alexander von Humboldt-Stiftung for a Feodor Lynen Postdoctoral Fellowship awarded to D.K.

Registry No. 2, 136675-81-5; **5a**, 136675-83-7; **6**, 135352-72-6; **7**, 136675-84-8; **8**, 136675-85-9; guanine, 73-40-5; 2-pivaloylguanine, 136675-82-6.

Direct, Highly Efficient Synthesis from (S)-(+)-Phenylglycine of the Taxol and Taxotere Side Chains

Jean-Noël Denis, Arlene Correa, and Andrew E. Greene*

Université Joseph Fourier de Grenoble, Chimie Recherche (LEDSS), Domaine Universitaire, BP 53X-38041 Grenoble Cedex, France

Received May 29, 1991

Taxol (3a, Chart I), a natural product obtainable in only low yield from yew bark,¹ and taxotere (3b), a semisynthetic analogue,² are very exciting antileukemic and tumor-inhibiting agents.³ Each of these substances, fortunately, can now⁴ be secured in good yield from the appropriate hydroxyl-protected side chain $(1a,b)^5$ and naturally abundant 10-desacetyl baccatin III (2, in protected form).⁶

The increasingly apparent cancer chemotherapeutic potential of these compounds has generated the need for a highly efficient enantioselective synthesis of the required side chains 1a,b. In this note we wish to disclose a particularly effective approach to these side chains from inexpensive, enantiomerically pure (S)-(+)-phenylglycine.

The synthetic strategy was based on the assumption that the aldehydes derived from alcohols 5a,b (Scheme I) would, under the proper conditions, undergo chelationcontrolled carbonyl addition⁷ and provide preferentially the desired threo amino alcohol derivatives 6a,b. It was, of course, presupposed that the aldehydes would have the necessary configurational stability for this approach.⁸

(3) "[Taxol] is one of the most promising new drugs in many years."
(National Cancer Institute request for applications for biological and chemical studies of taxol, July 27, 1990). Taxol (NSC-125973) and taxotree are currently undergoing clinical trials. See: Lomax, N. R.; Narayanan, V. L. Chemical Structures of Interest to the Division of Cancer Treatment; U.S. Government Printing Office: Washington, DC, 1983; Vol. III, p 17. Suffness, M.; Cordell, G. A. In The Alkaloids, Chemistry and Pharmacology; Brossi, A., Ed.; Academic Press: Orlando, FL, 1985; Vol. XXV, Chapter 1. Engel, S. L.; Schwartz, E. L.; Strauman, J. J.; Wiernik, P. H. Proc. Am. Assoc. Cancer Res. 1985, 26, 158. Zee-Cheng, R. K.-Y.; Cheng, C. C. Drugs Future 1986, 11, 45-48. Wiernik, P. H.; Schwartz, E. L.; Strauman, J. J.; Dutcher, J. P.; Lipton, R. B.; Paietta, E. Cancer Res. 1987, 47, 2486-2493. Rowinsky, E. R.; Cazenave, L. A.; Donehower, R. C. J. Natl. Cancer Inst. 1990, 82, 1247-1259.
(4) Denis, J.-N.; Greene, A. E.; Guénard, D.; Guéritte-Voegelein, F.;

(4) Denis, J.-N.; Greene, A. E.; Guénard, D.; Guéritte-Voegelein, F.; Mangatal, L.; Potier, P. J. Am. Chem. Soc. 1988, 110, 5917-5919. Recent improvements will be disclosed in due course.

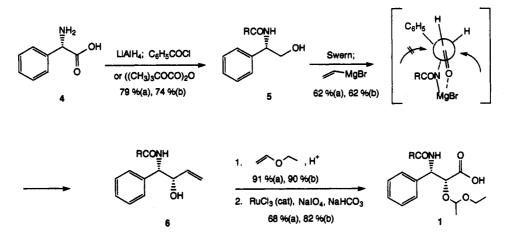
(5) (a) Denis, J.-N.; Greene, A. E.; Serra, A. A.; Luche, M.-J. J. Org. Chem. 1986, 51, 46-50.
(b) Denis, J.-N.; Correa, A.; Greene, A. E. Ibid.
1990, 55, 1957-1959. See also: Ojima, I.; Habus, I.; Zhao, M.; Georg, G. I.; Jayasinghe, L. R. Ibid. 1991, 56, 1681-1683.

(6) Chauvière, G.; Guénard, D.; Picot, F.; Sénilh, V.; Potier, P. C.R. Seances Acad. Sci., Ser. 2, 1981, 293, 501-503.

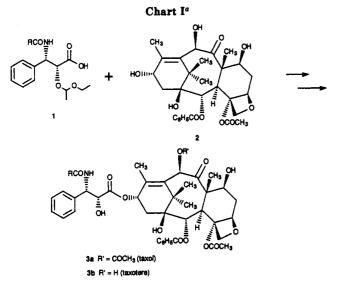
Seances Acad. Sci., Ser. 2, 1981, 293, 501-503.
(7) For related examples, see: Holladay, M. W.; Rich, D. H. Tetrahedron Lett. 1983, 24, 4401-4404. Hanson, G. J.; Lindberg, T. J. Org. Chem. 1985, 50, 5399-5401. Reetz, M. T.; Drewes, M. W.; Schmitz, A. Angew. Chem., Int. Ed. Engl. 1987, 26, 1141-1143. Thaisrivongs, S.; Pals, D. T.; Kroll, L. T.; Turner, S. R.; Han, F.-S. J. Med. Chem. 1987, 36, 76-982. Herold, P. Helv. Chim. Acta 1988, 51, 434-362. Garner, P.; Park, J. M.; Malecki, E. J. Org. Chem. 1988, 53, 4503-4508. Takemoto, Y.; Matsumoto, T.; Ito, Y.; Terashima, S. Tetrahedron Lett. 1990, 31, 217-218. Prasad, J. V. N. V.; Rich, D. H. Ibid. 1990, 31, 1803-1806. Polt, R.; Peterson, M. A. Ibid. 1990, 31, 4985-4986. Thompson, W. J.; Tucker, T. J.; Schwering, J. E.; Barnes, J. L. Ibid. 1990, 31, 6819-6822.

Wani, M. C.; Taylor, H. L.; Wall, M. E.; Coggon, P.; McPhail, A. T. J. Am. Chem. Soc. 1971, 93, 2325-2327. Miller, R. W.; Powell, R. G.; Smith, C. R.; Arnold, E.; Clardy, J. J. Org. Chem. 1981, 46, 1469-1474.
 Sénilh, V.; Blechert, S.; Colin, M; Guénard, D.; Picot, F.; Potier, P.; Varenne, P. J. Nat. Prod. 1984, 47, 131-137. Magri, N. F.; Kingston, D. G. I. J. Org. Chem. 1986, 51, 797-802.
 (2) Colin, M.; Guénard, D.; Guéritte-Voegelein, F.; Potier, P. Fr. Pat.

⁽²⁾ Colin, M.; Guénard, D.; Guéritte-Voegelein, F.; Potier, P. Fr. Pat. Appl. 86/10,400, 1986; Eur. Pat. Appl. EP 253,738, 1988. Mangatal, L.; Adeline, M. T.; Guénard, D.; Guéritte-Voegelein, F.; Potier, P. Tetrahedron 1989, 45, 4177-4190.



^aSeries a: $R = C_6H_5$. Series b: $R = (CH_3)_3CO$.



^aSeries a: $R = C_6H_5$. Series b: $R = (CH_3)_3CO$.

(S)-(+)-Phenylglycine (4) was conveniently and efficiently converted to the amino alcohol derivatives **5a**,**b** in a one-pot operation that involved lithium aluminum hydride reduction⁹ followed by in situ derivatization. The in situ treatment obviated the need to isolate the polar amino alcohol intermediate, and the yields of **5a**,**b** were thereby substantially improved.¹⁰

Although α -amino aldehyde derivatives are known to be configurationally labile,⁸ it was felt that the tandem Swern oxidation-carbonyl addition sequence developed by Ireland and Norbeck¹¹ might be employed successfully to effect a nonracemizing conversion of **5a,b** to **6a,b**. It was thus initially quite disheartening to find that the syn product that resulted from in situ treatment of the aldehyde from **5b** with excess vinylmagnesium bromide was essentially racemic. Reasoning that the initially produced carbamate anion, formed in the presence of unreacted aldehyde, might be responsible for this result, we added the Swern oxidation¹² mixture to the vinylmagnesium bromide in tetrahydrofuran-dichloromethane at room temperature. Much to our satisfaction, we found that the reaction, when carried out in this way, proceeded with good syn diastereoselection (9:1) and with complete retention of enantiomeric purity¹³ to give **6b** in 62% yield after purification. The pure amino alcohol derivative **6a** could be obtained in an analogous manner, also in 62% yield.

In anticipation of both the double bond oxidation and the ultimate esterification reaction (Chart I), the hydroxyls of **6a**,**b** were protected by reaction with ethyl vinyl ether in the presence of pyridinium *p*-toluenesulfonate.¹⁴ Of the several oxidation procedures examined, the combination of ruthenium chloride (catalytic) and sodium periodate¹⁵ in the presence of sodium bicarbonate^{5a} proved clearly the best and delivered pure **1a**,**b** in high yields. The enantiomeric purity (\geq 99%) of these substances was readily confirmed by ¹H and ¹⁹F NMR analysis of the Mosher esters¹³ of the alcohols derived from the methyl esters.

In summary, the pure hydroxyl-protected side chains of taxol and taxotere can now be easily obtained in only four steps from inexpensive (S)-(+)-phenylglycine, with overall yields of 30 and 34%, respectively. It is particularly significant that the approach is well-suited for large-scale work.

Experimental Section¹⁶

(-)-N-((S)-2-Hydroxy-1-phenylethyl)benzamide (5a). (S)-(+)-Phenylglycine (5.00 g, 33.1 mmol) was carefully added portionwise from the top of the condenser to a stirred mixture

(16) For general experimental procedures, see ref 5b.

⁽⁸⁾ For a discussion of the configurational stability of N-protected α -amino aldehydes, see: Jurczak, J.; Golebiowski, A. Chem. Rev. 1989, 89, 149–164. Phenyglycine derivatives are particularly racemization prone. See, for example: Burgess, K.; Ohlmeyer, M. J. Tetrahedron Lett. 1989, 30, 5857–5860. Evans, D. A.; Ellman, J. A.; DeVries, K. M. J. Am. Chem. Soc. 1989, 111, 8912–8914. It should be noted, however, that carbamate anion formation can render subsequent α -proton abstraction difficult, and thereby slow racemization. See: Coppola, G. M.; Schuster, H. F. Asymmetric Synthesis. Construction of Chiral Molecules using Amino Acids; Wiley Interscience: New York, 1987; Chapter 1, p 14. (9) Luly, J. R.; Dellaria, J. F.; Plattner, J. J.; Soderquist J. L.; Yi, N.

⁽⁹⁾ Luly, J. R.; Dellaria, J. F.; Plattner, J. J.; Soderquist J. L.; Yi, N. J. Org. Chem. 1987, 52, 1487-1492. Detectable racemization does not occur: Poindexter, G. S.; Meyers, A. I. Tetrahedron Lett. 1977, 3527-3528. Stanfield, C. F.; Parker, J. E.; Kanellis, P. J. Org. Chem. 1981, 46, 4799-4800.

⁽¹⁰⁾ For other examples, see: Correa, A.; Denis, J.-N.; Greene, A. E. Synth. Commun. 1991, 21, 1-9.

⁽¹¹⁾ Ireland, R. E.; Norbeck, D. W. J. Org. Chem. 1985, 50, 2198-2200.

⁽¹²⁾ For reviews, see: Mancuso, A. J.; Swern, D. Synthesis 1981, 165-185. Tidwell, T. T. *Ibid.* 1990, 857-870. Tidwell, T. T. Org. React. 1990, 39, 297-572. For an example of a large-scale (0.36 mol) Swern oxidation, see: Schore, N. E.; Knudsen, M. J. J. Org. Chem. 1987, 52, 569-580. See also: Taber, D. F.; Amedio, J. C., Jr.; Jung, K.-Y. *Ibid.* 1987, 52, 5621-5622.

⁽¹³⁾ Dale, J. A.; Dull, D. L.; Mosher, H. S. J. Org. Chem. 1969, 34, 2543-2549.

⁽¹⁴⁾ Miyashita, N.; Yoshikoshi, A.; Grieco, P. J. Org. Chem. 1977, 42, 3772-3774.

⁽¹⁵⁾ Carlsen, P. H. J.; Katsuki, T.; Martin, V. S.; Sharpless, K. B. J. Org. Chem. 1981, 46, 3936-3938.

of 2.52 g (66.3 mmol) of LiAlH₄ in 120 mL of tetrahydrofuran (THF) at reflux under argon. After the addition, the condenser was rinsed with 7 mL of THF, and the mixture was refluxed for an additional 6 h. The mixture was then allowed to cool to room temperature and was slowly treated with 4.0 mL of 10% aqueous NaOH followed by 5.0 mL of water and was stirred for 5 min. Additional 10% aqueous NaOH (53 mL) and 3.3 mL (4.0 g, 28 mmol) of benzoyl chloride were then introduced at 0 °C, and the resulting mixture was stirred for 30 min at 20 °C, whereupon CH₂Cl₂ and aqueous potassium sodium tartrate (Rochelle salt) were added. The crude product was isolated with CH₂Cl₂ in the usual way and recrystallized from CH₃OH-CH₂Cl₂ to give 4.90 g of 5a. Silica gel chromatography with 20% $CH_3CO_2C_2H_5$ in CH₂Cl₂ of the residue from the mother liquor yielded an additional 1.42 g (79%) of 5a: mp 179–180 °C (CH₂Cl₂-cyclohexane); $[\alpha]^{25}$ _D -18° (c 1.5, CH₃OH); ¹H NMR (300 MHz) δ 7.73-7.70 (m, 2 H), 7.42–7.16 (m, 8 H), 6.71 (br s, 1 H), 5.18 (dt, J = 4.8, 6.5 Hz, 1 H), 3.93 (dd, J = 4.8, 5.5 Hz, 2 H), 2.40 (t, J = 5.5 Hz, 1 H); ¹³C NMR (50.3 MHz, $CDCl_3 + CD_3OD$) δ 166.2, 139.2, 134.0, 131.6, 128.5 128.4, 127.5, 127.0, 126.6, 65.3, 55.8; IR 3300, 1630, 1580, 1520, 1075 cm⁻¹; mass spectrum (CI) m/z 242 (MH⁺), 224, 210, 122, 105. Anal. Calcd for C₁₅H₁₅O₂N: C, 74.66; H, 6.27; N, 5.81. Found: C, 74.45; H, 6.10; N, 5.91.

(+)-1,1-Dimethylethyl (N-((S)-2-Hydroxy-1-phenylethyl)amino)methanoate (5b). (S)-(+)-Phenylglycine (9.07 g, 60.0 mmol) was carefully added as above to 4.55 g (120 mmol) of LiAlH₄ in 210 mL of THF at reflux under argon. After the addition, the condenser was rinsed with 10 mL of THF, and the mixture was refluxed for an additional 6 h. The mixture was then allowed to cool to room temperature and was slowly treated with 7.3 mL of 10% aqueous NaOH followed by 9.1 mL of water and was stirred for 5 min. A solution of 14.40 g (66.0 mmol) of di-tert-butyl dicarbonate and 200 mg (1.64 mmol) of 4-(dimethylamino)pyridine in 80 mL of CH₂Cl₂ was introduced and the resulting mixture was refluxed for 6 h, whereupon it was allowed to cool to room temperature and was filtered through a pad of anhydrous Na₂SO₄, which was then washed with several portions of CH₂Cl₂. The solvents were removed under reduced pressure to give the crude product, which was recrystallized from CH_2Cl_2 -cyclohexane to give 7.50 g of 5b. Silica gel chromatography with 50% ether in hexane provided an additional 2.99 g (74%) of **5b**: mp 136–137 °C (CH₂Cl₂–cyclohexane); $[\alpha]^{24}_{D}$ +40° (c 1.6, CHCl₃); ¹H NMR (300 MHz) δ 7.38-7.26 (m, 5 H), 5.20 (br s, 1 H), 4.77 (br s, 1 H), 3.85 (d, J = 4.3 Hz, 2 H), 1.99 (br s, 1 H), 1.43 (s, 9 H); ¹³C NMR (50.3 MHz) δ 156.1, 139.6, 128.6, 127.6, 126.5, 79.9, 66.6, 56.7, 28.2; IR 3250, 1670, 1555, 1365, 1060 cm⁻¹; mass spectrum (CI) m/z 295 (M⁺ + isobutane), 255 (MH⁺ + ammonia), 238 (MH+, 100), 220, 206, 199, 182, 168, 150, 138, 124, 106. Anal. Calcd for C₁₃H₁₉O₃N: C, 65.80; H, 8.07; N, 5.90. Found: C, 65.89; H, 8.02; N, 5.76.

(-)-N-((1S,2S)-2-Hydroxy-1-phenyl-3-butenyl)benzamide (6a). To a stirred solution of 1.05 mL (1.52 g, 12.0 mmol) of oxalyl chloride in 16 mL of CH₂Cl₂ at -78 °C under argon was added 908 μ L (1.00 g, 12.8 mmol) of dimethyl sulfoxide (DMSO). After being stirred for 5 min at -78 °C, the reaction mixture was allowed to warm to -60 °C over 20 min, whereupon 1.88 g (7.79 mmol) of alcohol 5a suspended in 25 mL of CH₂Cl₂-DMSO (24:1) was added over 15 min. The flask containing the suspension was rinsed with 5 mL of CH₂Cl₂, which was then added to the reaction mixture. The mixture was allowed to warm to -35 °C over 20 min, stirred for 5 min at this temperature, and then treated over 4 min with 8.36 mL (6.20 g, 48.0 mmol) of diisopropylethylamine. The cooling bath was removed for 5 min, and then at -78 °C the mixture was added with a double-tipped needle to a room temperature solution (104 mL, 0.5 M, 52 mmol) of vinylmagnesium bromide in 1:1 THF-CH₂Cl₂ (exothermic!). After being stirred for 1 h, the mixture was treated with 8 mL of C_2H_5OH and 12 mL of saturated aqueous NH_4Cl . CH_2Cl_2 and aqueous HCl were then added and the crude reaction product was isolated with CH_2Cl_2 in the normal way. Purification of this material by silica gel chromatography with 5% CH₃CO₂C₂H₅ in CH₂Cl₂ gave 169 mg of 5a, 280 mg of anti product, and 1.17 g (62% based on consumed **5a**) of **6a**: mp 135–136 °C (CH₂Cl₂–cyclohexane); $[\alpha]^{23}_{D}$ -50° (c 1.0, CHCl₃); ¹H NMR (200 MHz) δ 7.83–7.80 (m, 2 H), 7.54-7.24 (m, 8 H), 6.98 (d, J = 7.6 Hz, 1 H), 5.94 (ddd, J = 5, 10.5, 17.1 Hz, 1 H), 5.40 (dt, J = 1.5, 17.1 Hz, 1 H), 5.26 (dd, J = 3.5, 7.6 Hz, 1 H), 5.23 (dt, J = 1.5, 10.5 Hz, 1 H), 4.55 (ddd, J = 3.5, 3.5, 5.0 Hz, 1 H), 2.40 (d, J = 3.9 Hz, 1 H); ¹³C NMR (50.3 MHz) δ 167.5, 139.6, 137.4, 134.3, 131.6, 128.7, 128.6, 127.7, 127.0, 126.9, 116.6, 75.3, 57.7; IR 3300, 1620, 1525, 1120, 1080, 995, 920 cm⁻¹; mass spectrum (CI) m/z 268 (MH⁺), 250, 210, 105. Anal. Calcd for C₁₇H₁₇O₂N: C, 76.38; H, 6.41. Found: C, 76.44; H, 6.49.

(+)-1,1-Dimethylethyl (N-((1S,2S)-2-Hydroxy-1-phenyl-3-butenyl)amino)methanoate (6b). To a reaction mixture prepared as above for 6a (but with 24 mL of CH₂Cl₂) was added over 15 min a solution of 1.90 g (8.01 mmol) of 5b in 26 mL of CH_2Cl_2 . The mixture was allowed to warm to -35 °C over 20 min, stirred for 5 min at this temperature, and then treated over 4 min with 8.36 mL (6.20 g, 48.0 mmol) of diisopropylethylamine. The reaction mixture was allowed to warm to -5 to 0 °C over 10 min and was then added with a double-tipped needle to a room temperature solution (104 mL, 0.5 M, 52 mmol) of vinylmagnesium bromide in 1:1 THF-CH₂Cl₂ (exothermic!). After being stirred for 1 h, the mixture was treated with 8 mL of C_2H_5OH and 12 mL of saturated aqueous NH₄Cl. CH₂Cl₂ and aqueous HCl were then added, and the crude reaction product was isolated with CH_2Cl_2 in the usual manner (ca. 87:13 syn-anti by ¹H NMR). Chromatography of this material on silica gel with 5% ether in CH₂Cl₂ gave 1.70 g (81%) of a 94:6 syn-anti mixture of products. Rechromatography of the mixture with 5% ether-45% CH₂Cl₂-50% hexane yielded 1.30 g (62%) of pure 6b: mp 56-57 °C; $[\alpha]^{25}_{D}$ +0.3° (c 1.6, CHCl₃); ¹H NMR (300 MHz) δ 7.37–7.24 (m, 5 H), 5.86 (ddd, J = 5.4, 10.5, 17.2 Hz, 1 H), 5.34 (dt, J = 1.4, J)17.2 Hz, 1 H), 5.26 (br s, 1 H), 5.20 (dt, J = 1.4, 10.5 Hz, 1 H), 4.70 (br s, 1 H), 4.38 (pseudo t, J = 4.6, 4.8 Hz, 1 H), 1.90 (br s, 1 H), 1.40 (s, 9 H); ¹³C NMR (50.3 MHz) δ 155.9, 140.0, 137.2, 128.3, 127.3, 126.7, 116.4, 79.6, 75.3, 58.7, 28.1; IR 3400, 1690, 1500, 1365, 1250, 1175, 1050, 920 cm⁻¹; mass spectrum (CI) m/z 321 (M⁺ + isobutane), 281 (MH⁺ + ammonia), 264 (MH⁺, 100), 246, 225, 208, 190, 164, 124, 106. Anal. Calcd for C₁₅H₂₁O₃N: C, 68.41; H, 8.04; N, 5.32. Found: C, 68.15; H, 7.98; N, 5.34

(2R,3S)-(-)-2-(1-Ethoxyethoxy)-3-phenyl-3-(phenylmethanamido)propanoic Acid (1a). A solution of 708 mg (2.65 mmol) of 6a and 66.5 mg (0.26 mmol) of pyridinium p-toluenesulfonate in 2.53 mL (1.91 g, 26.5 mmol) of ethyl vinyl ether and 13.5 mL of CH₂Cl₂ under argon was stirred at room temperature for 4 h. One drop of pyridine was added to the reaction mixture, which was then processed with CH_2Cl_2 in the normal manner. The crude product was purified by silica gel chromatography with 40% ether in hexane to provide 818 mg (91%) of N-((1S,2S)-2-(1ethoxyethoxy)-1-phenyl-3-butenyl)benzamide: mp 85.5-87 °C; $[\alpha]^{22}_{D}$ -34° (c 1.6, CHCl₃); ¹H NMR (200 MHz) δ 7.92-7.74 (m, 2 H), 7.55–7.15 (m, 8 H), 7.05 (d, J = 8 Hz, 1 H), 5.99 and 5.81 (2ddd, J = 6.5, 10.4, 17.1 Hz, 1 H), 5.46-5.12 (m, 3 H), 4.43-4.29(m, 1 H), 4.66–4.36 (2q, J = 5.3 Hz, 1 H), 3.56–2.98 (m, 2 H), 1.29 and 1.09 (2d, J = 5.3 Hz, 3 H), 1.08 and 1.00 (2t, J = 7.0 Hz, 3 H); IR 3300, 1630, 1600, 1580, 1520, 1125, 1030, 920 cm⁻¹; mass spectrum (CI) m/z 340 (MH⁺), 294, 268, 250, 211, 105.

To a stirred mixture of 254 mg (0.75 mmol) of the above acetal in 1.5 mL of CH₃CN, 1.5 mL of CCl₄, and 2.25 mL of H₂O at room temperature under argon were added 409.5 mg (4.88 mmol) of NaHCO₃ and, in small portions, 882 mg (4.12 mmol) of NaIO₄. After being stirred for 5 min following completion of the addition, the mixture was treated with 25.4 mg (0.12 mmol) of RuCl₃, and stirring was allowed to continue for 48 h. The reaction mixture was extracted with ether and then carefully acidified with aqueous HCl, and the product was isolated with CH_2Cl_2 to give 183 mg (68%) of pure 1a (as a ca. 1:1 mixture of methyl epimers): mp 93-94 °C; [α]²⁵_D -21° (c 0.7, CH₃OH); ¹H NMR (300 MHz) δ 7.80-7.00 (br s, 1 H), 7.83-7.78 (m, 2 H), 7.52-7.26 (m, 9 H), 5.80-5.74 (m, 1 H), 4.81 and 4.60 (2q, J = 5.3 Hz, 1 H), 4.63 and 4.50 (2d, J = 2.4 Hz, 1 H), 3.45–3.24 and 2.99–2.88 (2m, 2 H), 1.24 (d, J = 5.3 Hz, 3 H), 1.07 and 0.90 (2t, J = 7 Hz, 3 H); IR 3425, 3600-2100, 3060, 3025, 1740, 1640, 1600, 1580, 1520, 1480, 1075 cm^{-1} . The methyl ester of 1a (CH₂N₂) was identical with material previously prepared by an alternative synthesis.^{5a} ¹H and ¹⁹F NMR analysis of the Mosher esters of the alcohols derived from the methyl esters of (\pm) -la and (-)-la (aqueous HCl; (R)-(-)-2methoxy-2-phenyl-2-trifluoromethylacetyl chloride, pyridine) confirmed the enantiomeric purity $(\geq 99\%)$ of (-)-1a.

(2R, 3S)-(+)-3-(N-(1,1-Dimethylethoxycarbonyl)amino)-2-(1-ethoxyethoxy)-3-phenylpropanoic Acid (1b). A solution of 1.04 g (3.95 mmol) of 6b and 99 mg (0.40 mmol) of pyridinium p-toluenesulfonate in 3.80 mL (2.86 g, 39.7 mmol) of ethyl vinyl ether and 20 mL of CH₂Cl₂ under argon was stirred at room temperature for 4 h. One drop of pyridine was added to the reaction mixture, which was then processed with CH_2Cl_2 in the usual way. The crude product was purified by silica gel chromatography with 20% ether in hexane to provide 1.19 g (90%) of 1,1-dimethylethyl (N-((1S,2S)-2-(1-ethoxyethoxy)-1-phenyl-3-butenyl)amino)methanoate: mp 59–65 °C; $[\alpha]^{25}_{D}$ +15° (c 1.6, CHCl₃); ¹H NMR (300 MHz) & 7.37–7.17 (m, 5 H), 5.91 and 5.77 (2ddd, J = 7, 10.5, 17.4 Hz, 1 H), 5.44 and 5.37 (2m, 1 H), 5.30and 5.25 (2dt, J = 1.2, 17.4 Hz, 1 H), 5.23 and 5.22 (2dt, J = 1.2, 10.5 Hz, 1 H), 4.73 and 4.71 (2m, 1 H), 4.62 and 4.31 (2q, J = 5.4and 5.3 Hz, 1 H), 4.23 and 4.16 (2 pseudo dd, J = 6.6, 7.0 Hz, 1 H), 3.51-3.05 and 2.98-2.90 (2 m, 2 H), 1.40 (s, 9 H), 1.22 and 1.05 (2d, J = 5.4 and 5.3 Hz, 3 H), 1.07 and 0.90 (2t, J = 7.0 Hz, 3 H);IR 3370, 1680, 1520, 1170, 1080, 1050 cm⁻¹. Anal. Calcd for C19H29O4N: C, 68.03; H, 8.71; N, 4.18. Found: C, 68.00; H, 8.78;

N, 4.13. The above acetal (1.09 g, 3.25 mmol) was treated as was that from 6a to afford 940 mg (82%) of pure 1b: mp 33-37 °C; $[\alpha]^{25}$ +18° (c 1.1, CHCl₃); ¹H NMR (300 MHz) δ 8.52 (br s, 1 H), 7.38-7.13 (m, 5 H), 5.72 (br s, 1 H), 5.29 (br s, 1 H), 4.80-4.65 and 4.50-4.35 (2m, 2 H), 3.52-3.15 and 2.88-2.60 (2m, 2 H), 1.42 (s, 9 H), 1.20 and 1.18 (2d, J = 5.4 Hz, 3 H), 1.04 and 0.81 (2t, J = 7.0 Hz, 3 H); IR 3700-2200, 3060, 1720, 1660, 1370, 1170, 1080, 955 cm⁻¹. The methyl ester of 1b (CH_2N_2) was identical with material previously prepared by an alternative synthesis.^{5b} ¹H and ¹⁹F NMR analysis of the Mosher esters of the alcohols derived from the methyl esters of (\pm) -1b and (+)-1b (aqueous HCl; (R)-(-)-2-methoxy-2-phenyl-2-trifluoromethylacetyl chloride, pyridine) confirmed the enantiomeric purity $(\geq 99\%)$ of (+)-1b.

Acknowledgment. We thank Prof. J. Lhomme and Dr. J.-L. Luche for their interest in our work and Dr. D. Grierson for useful suggestions. Financial support from the CNRS (URA 332) and Rhône-Poulenc Rorer and a fellowship award from the CAPES (Brazil) to A.C. are gratefully acknowledged.

Registry No. 1a (isomer 1), 136778-67-1; 1a (isomer 2), 136778-69-3; 1a (methyl ester; isomer 1), 136779-75-4; 1a (methyl ester; isomer 2), 136778-73-9; 1a (methyl ester; Mosher ester), 136693-08-8; 1b (isomer 1), 136778-68-2; 1b (isomer 2), 136778-70-6; 1b (methyl ester; isomer 1), 136778-71-7; 1b (methyl ester; isomer 2), 136778-75-1; 1b (methyl ester; Mosher ester), 136693-09-9; 2, 32981-86-5; 3a, 33069-62-4; 3b, 114977-28-5; 4, 2935-35-5; 5a, 116126-04-6; 5b, 117049-14-6; 6a, 136693-02-2; anti-6a, 136693-06-6; 6a ((R)-ethoxyethyl ether), 136693-04-4; 6a ((S)-ethoxyethyl ether), 136778-72-8; 6b, 136693-03-3; anti-6b, 136693-07-7; 6b ((R)-ethoxyethyl ether), 136778-74-0; **6b** ((S)-ethoxyethyl ether), 136693-05-5.

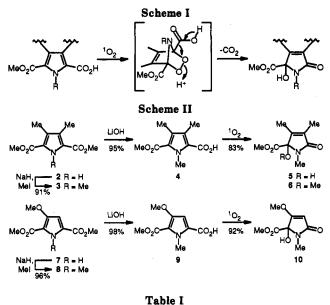
Singlet Oxygen Mediated Oxidative Decarboxylation of Pyrrole-2-carboxylic Acids

Dale L. Boger* and Carmen M. Baldino

Department of Chemistry, Purdue University, West Lafayette, Indiana 47907

Received June 6, 1991

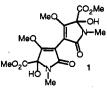
Autoxidation of the chromogen hermidin¹ isolated from Mercurialis perennis L.² provides isochrysohermidin (1),³ a functionalized 3,3'-pyrrolin-2-one dimer first isolated from Mercurialis leiocarpia and whose structure was unambiguously established by X-ray crystallography. In initial studies directed at the total synthesis of iso-



pyrrole	solvent (0.8 mM) ^a	time (h)	result
4	CH ₃ OH ^b	5	5 (20%), 6 (12%)
	CH ₃ OH	1	5 (35%), 6 (32%)
	CH ₃ OH-H ₂ O (2:1)	1	5 (37%), 6 (26%)
	ⁱ PrŎH-H ₂ Ō (2:1)	1	5 (79%)
	$^{i}PrOH-H_{2}O$ (3:1)	1	5 (83%)
	$CH_{3}CN - H_{2}O(3:1)$	1	5 (62%)
9	$CH_{3}CN-H_{2}O(3:1)$	1	10 (92%)
	ⁱ PrŎH–H ₂ Ō (3:1)	1	10 (80%)
	-		

^aRose bengal (8 mequiv), quartz immersion well, Hanovia highpressure mercury lamp (450 W), uranium yellow glass filter (transmits > 330 nm), O_2 , 22 °C. ^bPyrex reaction vessel, tungsten lamp (500 W), O_2 , 22 °C.

chrysohermidin,⁴ we have examined the singlet oxygen $({}^{1}O_{2})$ addition⁵ to substituted 5-(alkoxycarbonyl)pyrrole-2-carboxylic acids and a subsequent oxidative decarboxylation⁶ reaction in efforts to provide direct access to the 5-(alkoxycarbonyl)-5-hydroxy-3-pyrrolin-2-one subunit found in 1, Scheme I.



The substrates employed in the study were derived from the [4 + 2] cycloaddition of 2-[(triethylsilyl)oxy]-2-butene and 1,1-dimethoxyethylene with 3,6-bis(methoxycarbonyl)-1,2,4,5-tetrazine7 followed by reductive ring contraction (Zn, HOAc) of the resulting 1,2-diazine cycloadducts to provide the substituted pyrroles 2 and 7.8 N-Methylation of the pyrroles followed by a surprisingly selective monohydrolysis of the symmetrical pyrrole 3 and a well-precedented⁹ selective hydrolysis of the sterically

- Swan, G. A. Experientia 1984, 40, 687.
 Haas, P.; Hill, T. G. Biochem. J. 1925, 19, 233.
 Masui, Y.; Kawabe, C.; Mastumoto, K.; Abe, K.; Miwa, T. Phytochemistry 1986, 25, 1470.

^{*}Address correspondence to this author at: Department of Chemistry, The Scripps Research Institute, 10666 N. Torrey Pines Road, La Jolla, CA 92037.

⁽⁴⁾ Abe, K.; Okada, T.; Masui, Y.; Miwa, T. Phytochemistry 1989, 28, 960.

⁽⁵⁾ Lightner, D. A.; Quistad, G. B. J. Heterocycl. Chem. 1973, 10, 273.
Lightner, D. A.; Crandall, D. C. Experientia 1973, 29, 262.
(6) Jefford, C. W.; Boschung, A. F.; Bolsman, T. A. B. M.; Moriarty, R. M.; Melnick, B. J. Am. Chem. Soc. 1976, 98, 1017. Schenck, G. O.

Liebigs Ann. Chem. 1953, 584, 156. (7) Boger, D. L.; Panek, J. S.; Coleman, R. S.; Sauer, J.; Huber, F. X. J. Org. Chem. 1985, 50, 5377.

⁽⁸⁾ Boger, D. L.; Coleman, R. S.; Panek, J. S.; Yohannes, D. J. Org. Chem. 1984, 49, 4405.